PCBs as Environmental Estrogens: Turtle Sex Determination as a Biomarker of Environmental Contamination

Judith M. Bergeron, David Crews, and John A. McLachlan

¹Department of Zoology, Institute of Reproductive Biology, University of Texas at Austin, Austin, TX 78712 USA; ²National Institute of Environmental Health Sciences, Research Triangle Park, NC 27709 USA

Polychlorinated biphenyls (PCBs) are widespread, low-level environmental pollutants associated with adverse health effects such as immune suppression and teratogenicity. There is increasing evidence that some PCB compounds are capable of disrupting reproductive and endocrine function in fish, birds, and mammals, including humans, particularly during development. Research on the mechanism through which these compounds act to alter reproductive function indicates estrogenic activity, whereby the compounds may be altering sexual differentiation. Here we demonstrate the estrogenic effect of some PCBs by reversing gonadal sex in a reptile species that exhibits temperature-dependent sex determination. Key words: biomarker, environmental estrogens, gonadal sex, PCBs, sexual differentiation, temperature-dependent sex determination. Environ Health Perspect 102:780-781 (1994)

Many polychlorinated biphenyls (PCBs) are industrialized chemicals such as those used in adhesives, fire retardants, and waxes (1). As a function of physical and chemical properties such as lipid solubility and low rate of degradation, PCBs persist in the environment; thus, individuals in industrialized nations are exposed to high levels of these compounds (2,3). Because chemicals of this nature tend to bioaccumulate, for instance, within food chains, they eventually reach measurable levels in human tissues or in nutrient systems such as placental cord blood, breast milk, and egg yolks. This may result in an individual being exposed to high levels during development, a time when toxic effects may be more detrimental than those seen in adults (3,4).

Reproductive disorders resulting from exposure to xenobiotic estrogens may include reductions in fertility, lower hatch rates in fish and birds, and decreased viability of offspring, as well as alterations in hormone levels or adult sexual behaviors, all of which have further implications, particularly in wildlife population dynamics (3–5). In addition, there is increasing suspicion that effects of estrogenic compounds are correlated to disorders of the male reproductive system, including increased occurrence of prostatic and testicular cancers (6,7). There is a need for a sensitive bioassay with which the developmental effects of environmental

estrogens can be determined. Reptiles with temperature-dependent sex determination (TSD), in which the incubation temperature of the egg determines the sex of the individual, may provide such a bioassay. Evidence of PCBs acting as estrogens in a TSD species may contribute to a clearer understanding of how estrogens may lead to reproductive dysfunction.

The red-eared slider turtle, Trachemys scripta, is a species that possesses a TSD pattern in which warm incubation temperatures, (e.g., 31°C) produce all female hatchlings, whereas cooler incubation temperatures (e.g., 26°C) produce all male hatchlings; intermediate temperatures (between 29.0 and 30.0°C) result in varying ratios of males and females (8,9). In reptiles with TSD, the actions of estrogen mimic those of temperature with regard to gonadal differentiation (10,11). In the redeared slider, as in many other TSD species, exogenous estrogens applied to the eggshell during the period of sexual differentiation can counteract the effects of male-producing temperatures and induce ovarian development (12-15).

Materials and Methods

Eggs for these experiments were purchased from Robert Kliebert (Hammond, Louisiana) and incubated on a layer of vermiculite: water (1:1) in temperature-controlled chambers (27.8 or 26°C). We monitored embryonic development by candling eggs during early stages, then by dissecting a small sample of eggs approximately twice a week to determine specific developmental stages. At the beginning of the period of gonadal differentiation (stage 17: approximately 4 weeks from the date eggs are laid), which coincides with the developmental stage at which the embryos are sensitive to the effects of exogenous estradiol (10,11), eggs were randomly assigned to treatment groups and spotted with either PCB compounds (Ultra Scientific, Hope, Rhode Island) in 95% ethanol, estradiol-17ß (Sigma, St. Louis, Missouri) in ethanol as a positive (hormone) control, or with ethanol alone as a negative (temperature) control. Incubation was continued at the experimental temperatures until hatch (approximately 7 more weeks at this temperature), at which time we dissected the hatchlings to determine resulting sex ratios. We determined gonadal sex and status of genital ducts by visualization under a dissection microscope and verified sex histologically, as described previously for this species (8,9). In all cases that the gonad was female, the oviducts or Müllerian ducts were present. In some cases in which testes developed, the Müllerian ducts were also present, indicating estrogenic activity.

Results and Discussion

In the first experiment, 11 different PCB compounds believed to be estrogenic (16) were applied individually to eggs incubated at 27.8°C. As shown in Table 1, each compound was administered at 2 doses, 15 eggs per dose. The eggs average 11.4 g (±0.21) each. Comparison of the resulting sex ratios showed that two compounds, 2',4',6'-trichloro-4-biphenylol (compound F) and 2',3',4',5'-tetrachloro-4-biphenylol (compound G), significantly reversed sex at a male-producing temperature (Fisher's Exact Test, p < 0.001). The former compound showed 100% sex reversal at 100 μg, or just below 9 ppm. In tests using mouse tissue, these same two compounds showed an appreciable affinity for estrogen receptor, due in part to their conformational properties as hydroxybiphenyls (16,17). As metabolites of other PCBs, hydroxylated PCBs such as F and G may exist in steady-state concentrations in aquatic environments, potentially exposing wildlife to their effects via direct contact or through the food chain (17).

Because purified PCB compounds are rarely found in the environment, we decided in the second series of experiments to look at combinations of the same PCBs. All eggs were incubated at 27.8°C and received a low (10 µg), medium (100 mg), or high (145–190 µg) dose of compounds in ethanol. Some eggs received a cocktail of all PCBs except the two that caused sex reversal (F and G). Others were exposed to combined hydroxybiphenyls, excluding F and G. Lastly, some eggs were spotted with combined nonhydroxylated PCBs. In all three conditions, there was no evidence of sex reversal.

Address correspondence to J. M. Bergeron, Department of Zoology, Institute of Reproductive Biology, University of Texas at Austin, Austin, TX 78712 USA.

We acknowledge the technical assistance of A. Alexander, T. Cole, H. Nguyen, and J. O'Brien, and J. Branch for graphics. This research was supported by a National Science Foundation predoctoral fellowship to J.M.B. and National Institute of Mental Health Research Scientist Award 00135 to D.C.

Received 10 March 1994; accepted 27 June 1994.

Table 1. Effects of some PCB compounds on sex determination^a

Compound	% Hatchlings with female gonads (low dose/high dose)	% Hatchlings with oviducts (low dose/high dose)
A 2',5',-dichloro-3-biphenylol	0/0	0/14
B 2,2',4-trichlorodiphenyl ether	7/0	21/0
C 2,2',5'-trichloro-4-biphenylol	0/0	7/0
D 2,3,4'-trichlorobiphenyl	7/0	0/0
E 2,3',5-trichlorobiphenyl	0/8	7/8
F 2',4',6'-trichloro-4-biphenylol	0/100	0/100
G 2',3',4',5'-tetrachloro-4-biphenylol	4/50	8/71
H 2,4,4',5-tetrachlorodiphenyl ether	0/0	0/0
J 2,4,4',6-tetrachlorobiphenyl	7/0	0/7
K 2,4,4",6-tetrachloro-p-terphenyl	0/0	0/7
L 2',3',4',5,5'-pentachloro-2-biphenylol	0/0	0/0
Ethanol control	0	0
Estradiol-17ß control	100	100

^aEleven compounds were applied to eggs incubated at 27.8°C in two doses per compound. The doses were: A, B, C, F, G, H, J = 10 μ g, 100 μ g; D, L = 5 μ g, 50 μ g; E = 25 μ g, 250 μ g; K = 3.35 μ g, 33.5 μ g. The estradiol control consisted of 10 μ g estradiol-17 β s.

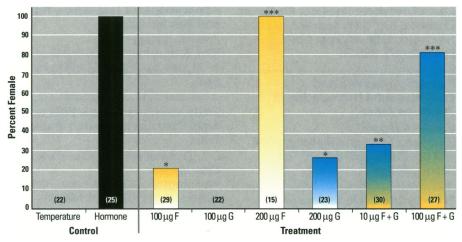


Figure 1. Percentage of female hatchlings at a temperature that normally produces 100% males (26°C) after treatment with two estrogenic PCB compounds in ethanol. Controls: temperature, ethanol alone; hormone, 10 μ g estradiol-17 β . PCB compounds: F, 2',4',6'-trichloro-4-biphenylol; G, 2',3',4',5'-tetrachloro-4-biphenylol. Sample sizes in parentheses. Significant sex reversal is indicated by * $p \le 0.03$; ***p = 0.003; ***p = 0.0001.

Because we knew compounds F and G showed estrogenic activity at the slightly higher temperature, we decided to test these two compounds at a temperature that produces 100% males (26°C). Both compounds showed significant sex reversal at this temperature (Fig. 1). When combined, F and G synergized, resulting in a significant increase in ovarian development at a dose of 10 µg, or less than 1 ppm (Fisher's Exact Test, p < 0.01), whereas F alone and G alone required at least a 10fold higher dose to show sex reversal. Exogenous estradiol-17ß produces similar results at a dose of 0.5 µg, or .04 ppm (9). As with the earlier experiments involving compounds F and G, whenever the gonad was feminized, the oviducts were retained, and in some cases where testes developed, there were also Müllerian ducts present.

The nature of TSD in this and other reptile species provides a useful system in which to assay the extent of estrogenic activity found in xenobiotic compounds. This report contributes laboratory evidence of the effect of PCBs on sex determination, emphasizing the usefulness of a TSD species as a biomarker to assess environmental contamination and serve as a warning of conditions threatening wildlife populations. The PCB levels reported here as effective in disrupting normal gonadal differentiation in the turtles are comparable to average levels of PCBs found in human breast milk in industrialized nations (3). In addition, this study supports the contention that environmental estrogens, through their action on reproductive development, have the potential to alter wildlife populations as well as contribute to reproductive dysfunction in humans (3-7). Future studies investigating the mechanisms through which these estrogenic compounds act to affect sex differentiation will continue to shed light on a human contribution to the environment.

REFERENCES

- Safe SH. Polychlorinated biphenyls (PCBs), dibenzo-p-dioxins (PCDDs), dibenzofurans (PCDFs), and related compounds: environmental and mechanistic considerations which support the development of toxic equivalency factors (TEFs). Crit Rev Toxicol 21:51-88 (1990).
- Lione A. Polychlorinated biphenyls and reproduction. Reprod Toxicol 2:83–89 (1988).
- Thomas KB, Colborn T. Organochlorine endocrine disruptors in human tissue. In: Chemically induced alterations in sexual and functional development: the wildlife-human connection (Colborn T, Clement C, eds). Princeton, NJ: Princeton Scientific, 1992; 365-394.
- Bern HA. The fragile fetus. In: Chemically induced alterations in sexual and functional development: the wildlife human connection (Colborn T, Clement C, eds). Princeton, NJ: Princeton Scientific, 1992;9–15.
- Gray LE, Jr. Chemical-induced alterations of sexual differentiation: a review of effects in humans and rodents. In: Chemically induced alterations in sexual and functional development: the wildlife-human connection (Colborn T, Clement C, eds). Princeton, NJ:Princeton Scientific, 1992;203–230.
- Sharpe RM, Skakkebaek NE. Are oestrogens involved in falling sperm count and disorders of the male reproductive tract? Lancet 341:1392–1395 (1993).
- Santti R, Newbold RR, Makela S, Pylkkanen L, McLachlan JA. Development estrongenization and prostatic neoplasia. Prostate 24:67–78 (1994).
- 8. Wibbels T, Bull JJ, Crews D. Chronology and morphology of temperature-dependent sex determination. J Exp Zool 260:371-381 (1991).
- 9. Wibbels T, Bull JJ, Crews D. Synergism between temperature and estradiol: a common pathway. J Exp Zool 260:130–134 (1991).
- Reynaud A, Pieau C. Embryonic development of the genital system. In: Biology of the reptilia, vol 15 (Gans C, Billet F, eds). New York: Wiley, 1985:149–300.
- 11. Wibbels T, Gideon P, Bull JJ, Crews D. Estrogen- and temperature-induced medullary regression during gonadal differentiation in a turtle. Differentiation 53:149–154 (1993).
- 12. Crews D, Bull JJ, Wibbels T. Estrogen and sex-reversal in turtles: dosages producing both sexes produce few intersexes. Gen Comp Endocrinol 81:357–364 (1991).
- Wibbels T, Bull JJ, Crews D. Hormoneinduced sex determination in an amniotic vertebrate. J Exp Zool 262:454

 –457 (1992).
- Bull JJ, Guizke WHN, Crews D. Sex reversal by estradiol in three reptilian orders. Gen Comp Endocrinol 70:425–428 (1988).
- Wibbels T, Crews D. Specificity of steroid hormone-induced sex determination in a turtle. J Endocrinol 133:121–129 (1992).
- Korach KS, Sarver P, Chae K, McLachlan JA, McKinney JD. Estrogen receptor-binding activity of polychlorinated hydroxybiphenyls: conformationally restricted structural probes. Mol Pharmacol 33:120–126 (1988).
- 17. McKinney JD, Korach KS, McLachlan, JA. Detoxification of polychlorinated biphenyls. Lancet 335:222-223 (1990).